

ANDREW GOREA:

EXHIBIT
1

BBMM

RECEIVED 60 CC CORD BLOOD (w/ CPD)
 PEC mons 30%
 T cells 70%

added 15 ml 1xPBS to bring to incubation
 volume of 45ml.

Added 1.5 ml antibody (12.8)
 incubated 25ml.

Primed CellPro "celfrate".

Spin cells. Resuspended in 1xPBS to
 a final volume of 300cc in bag.

Ran through column.

Unabsorbed portion → spun down.
 and consolidated in 1xPBS
 for incubation.

75ml for incubation (added HSA)
 1.5ml antibody (12.8) 25ml. Spin down
 following incubation. Resp to Vd. of 300cc in bag
 Ran through 2nd column.

Sel'm cell portions from Runs 1 & 2
 were combined (after counts done and
 samples removed for staining)

Total cells 2.8×10^6 for transduction

1L3 SANDE
 WEN
 1L6 SANDE
 WEN
 SCF WEN
 ANTRG
 WEN

Final concn
 will be dil
 Cells are
 concentrate

BBMM: FB
 BSO

2.8×10^6 cells
 P+ M

BBMM + 31b15CF (for 500ml of media)

1L3 SANDBZ * Y0230392 stock at 150ug/ml
Want final: 20ng/ml x 2 .. 20ug 133ul add
1L6 SANDBZ * Y0450392 stock at 150ug/ml
Want final: 50ng/ml x 2 .. 50ug 333ul add
SLF ABGEN * 1509F2 stock at 1.5mg/ml = 1500ug/ml
Want final: 100ng/ml x 2 .. 100ug 67ul add

Final concentrations are doubled since the media
will be diluted 1:2 w/ viral supernatant.
Cells are therefore incubated with the correct
concentrations.

BBMM: FBS Granuni lot# 940003H
BSA #115

2.8x10⁶ cells want final: 5x10⁴ cells.
Put in 2 T-75 30ml each: 15ml B365 051193
15ml LASU^{G7} lot# 53
+ protamine sulfate, 24ml
of 1:10 diluted stock

run down
P 300ml in bag.

42
end

Cord Blood cells pre processing:

CFUs

set 143

Start:

Plate #	Sample	# Cells	# uL/ml media
- G418	1ab	5×10^4	50
+ G418	2ab		50
- G418	3ab		100
+ G418	4ab	1×10^5	100

adsorb fraction:

CFUs Post transduction: set 144

Plate #	# Cells	# cell
0 G418	1ab	7
	2ab	14
	3ab	28
+ G418	4ab	7
	5ab	14
	6ab	28

(yields) adsorb

COUNT:

$$\bar{x} = 34$$

$$\times 2 \times 10^4 = 6.8 \times 10^5 \text{ Clones}$$

$$\times 5.5 \text{ mL} = 3.7 \times 10^{10} \text{ C}$$

X-enfused on 5/15/93
posttransduced stem cells

G418

+ -

7ab
8ab1000
200020
40adsorb
yields from
clone

Start:

$$5 \times 10^8 \text{ C}$$

PRE

$$0.71^\circ \text{C}$$

pre ab

$$0.22^\circ \text{C}$$

ml/ml media50
20
00
100

set 144

$$*34+ = 3.6 \times 10^8 \text{ C} = 1.1 \times 10^8 \text{ C}$$

adsorbed
fraction #1:

$$2 \times 10^8 \text{ C}$$

FL1 FL2 gate

$$31.94\%$$

FL1 FL2 gate

$$20.81\%$$

$$*34+ = 0.64 \times 10^8 \text{ C}$$

$$= 0.42 \times 10^8 \text{ C}$$

adsorbed
fraction #2:

$$0.8 \times 10^8 \text{ C}$$

$$2.4 \times 10^8 \text{ C}$$

$$5.80\%$$

$$*34+ c = 0.02 \times 10^8 \text{ C}$$

$$0.05 \times 10^8 \text{ C}$$

(yields)

adsorbed #1:

PRE & FL1 FL2 gate

$$\frac{0.64 \times 10^8 \text{ C}}{3.6 \times 10^8 \text{ C}} = 17.8\%$$

PRE & FL1 FL2 gate

$$\frac{0.42 \times 10^8 \text{ C}}{3.6 \times 10^8 \text{ C}} = 11.7\%$$

pre ab & FL1 FL2 gate

$$\frac{0.64 \times 10^8 \text{ C}}{1.1 \times 10^8 \text{ C}} = 58.2\%$$

Post ab & FL1 FL2 gate

$$\frac{0.42 \times 10^8 \text{ C}}{1.1 \times 10^8 \text{ C}} = 38.2\%$$

adsorbed #2:

yields from
bottom

PRE & FL1 FL2

inhibition

PRE & FL1 FL2

ZACHARY BIGGINS:

5/14/93

RECD 200cc cold blood

PRE: mono 109 poly 109
 Y

$$218 \times 50 = 10.9 \times 10^6 \text{ cells}$$

$$\times 200\text{ml} = 2.2 \times 10^9 \text{ C}$$

start

Added 3 vials (4.5ml) 12.8 ab.
 inc. 25 min.

Spindown. Rspd. in 1xPBS to 300ml
 in bag.

Ran through column.

Spindown unabsorbed fraction for 2nd ab
 incubation.

Spindown stem cell fraction 40 Rspd. in
 smaller volume for count.

Counts:

unabsorbed

$$\text{mono} \quad \text{poly}$$

$$67 \quad 102$$

$$10.9 \times 50 \times 10^3$$

$$= 8.5 \times 10^6 \text{ cells} \times 225\text{ml}$$

$$= 1.9 \times 10^9 \text{ C}$$

stem

$$\text{mono} \quad \text{poly}$$

$$172 \quad 16$$

$$188 \times 2 \times 10^4$$

$$= 3.8 \times 10^6 \text{ cells} \times 5.5\text{ml}$$

$$= 20.7 \times 10^6 \text{ C}$$

inc
 12.8
 300
 Rspd
 Rsp

Cuv

1

2

3

3.

=

Per
 free

com

26x1
 wan

= 2

13 fl.

LASI

Incubated unadsorbed fraction w/ 4.5 ml
2.8 ab. for 25 min.

Spun down.

Rot in 300ml in bag (w/ 1x PBS)
Ran through 2nd column.

Counts:

unadsorbed

monos polys

30 ✓ 33

$6.3 \times 5.3 \times 10^3$

$$3.15 \times 10^4 \text{ Cl/ml} \times 600 \text{ ml} \\ = 1.9 \times 10^6 \text{ C}$$

↓

stem

monos polys

58 ✓ 4

$6.2 \times 2 \times 10^4$

$$= 1.2 \times 10^6 \text{ Cl/ml}$$

$$\times 5 \text{ ml} = 6 \times 10^6 \text{ C}$$

start

20ml

Speduled/ficoll'd
froze \Rightarrow LWT(2)

combined stem cell fractions

$2.6 \times 10^6 \text{ C}$ for transduction

want final $[C] = 5 \times 10^4 \text{ Cl/ml}$

520 ml total

$= 2 = 200 \text{ ml sup}$

260 ml media

13 flasks 40 ml/flask

30 ml sup

20 ml media (B365)

+ 300 ml protamine sulfate

45

LASN sup 539 (bottles 18/19)

$\times 10^4$

ml $\times 5.5 \text{ ml}$

CFUs:

50745

PRE

Plate #	Example	# Cells	# ill
1ab	(-G418) PRE trans ↓ (+G418)	5,000,000 ↓	5
2ab			5

BBMM + 3161SCF:

1L3 Sandoz * y0230392

1L6 Sandoz * y04150392

SCF AMGEN * 150953

Took sample to micro for sterility ✓
 each day of transduction pd.
 Stat Gram stain done (negative)
 before cells were given to baby.

5/15/93 4pm 2nd transduction:

Spun cells down from each flask
 Respd in fresh media & LASN supe
 added Prostaglandin sulfate

5/16/93 3rd transduction 330pm.
 Repeated above.

5/17

CCR

PDT

Reir

Post

OG

+G4

OGHSE

+G4

5/17/93 cells washed 2x
 3x in 1x PBS + FBS
 last wash in RPMI (no FBS)

count: 60×10^6 c

$$\bar{x} = 15 \times 10 \times 10^6 = 1.5 \times 10^8$$

$$\times 40 \text{ ml} = 60 \times 10^6 \text{ c}$$

5
5

Put in 5cc into 10cc supernatant

Reinfused on 5/17/93 (UCSF)

Post trans. CFUs: 50146

Sample/plate #	# cells	# el
06418 1ab	500	4
06418 2ab	1000	8
06418 3ab	2000	16
06418 4ab	500	4
06418 5ab	1000	8
06418 6ab	2000	16
06418 7ab	1000	24
06418 8ab	1000	24

✓

(4/16)

> baby

tion:

in flask

SN supe

> 30pm